

1. Hydrolytic activity of lipase on *p*-nitrophenyl ester (Kademi *et al.*, 2000)

Hydrolytic activity of lipase solution was carried out by spectrophotometric method using *p*-nitrophenyl ester as substrate. The concentration of *p*-nitrophenol was measured at 410 nm. One unit of enzyme was the amount of enzyme liberating one μmol *p*-nitrophenol/mL/min under the determined conditions. The concentration of *p*-nitrophenol was calculated by Lambert and Beer equation:

$$A = \frac{\Delta E}{\epsilon \times d \times c}$$

Where	A	=	Activity (U/mL)
	ΔE	=	Absorbance at 410 nm
	ϵ	=	Molar extinction coefficient (L/mol/cm)
	d	=	Cuvette width (cm)
	c	=	Amount of enzyme (mL)

The molar extinction coefficient (ϵ) of *p*-nitrophenol was depended on pH value (Figure 69).

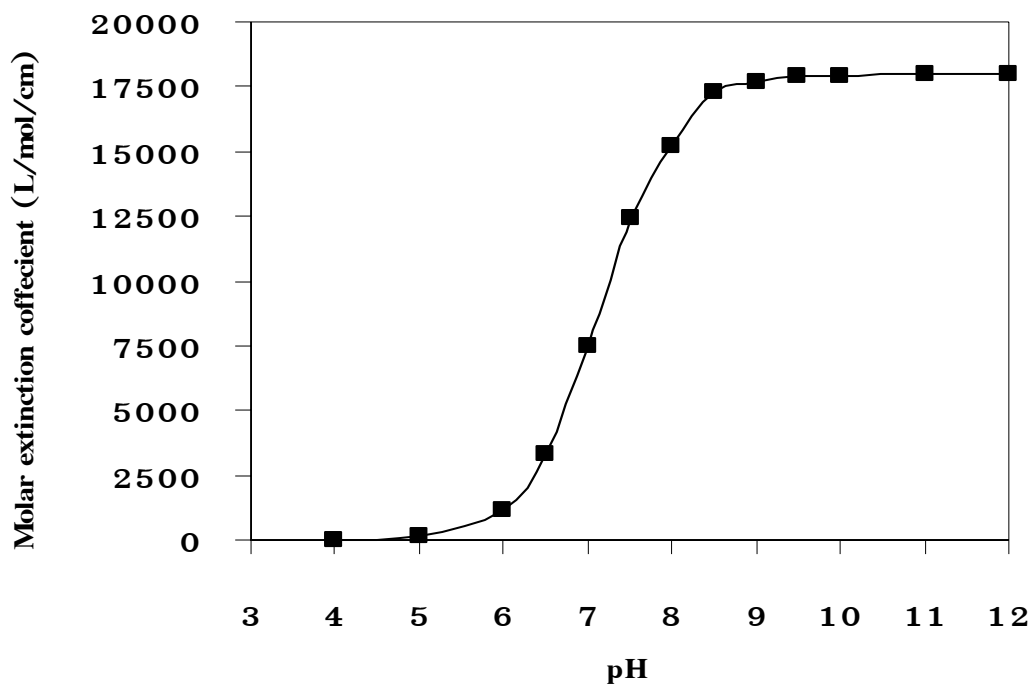


Figure 69. Calibrating curve of molar extinction coefficient of *p*-nitrophenol with different pH.

2. Hydrolytic activity of lipase by cupric acetate method (Lee and Rhee, 1993)

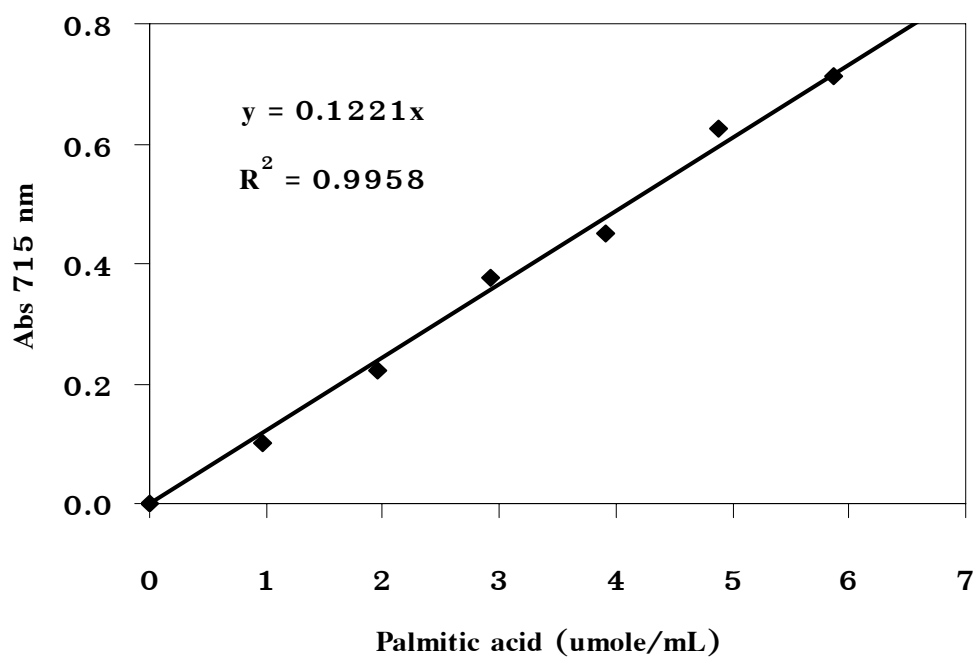
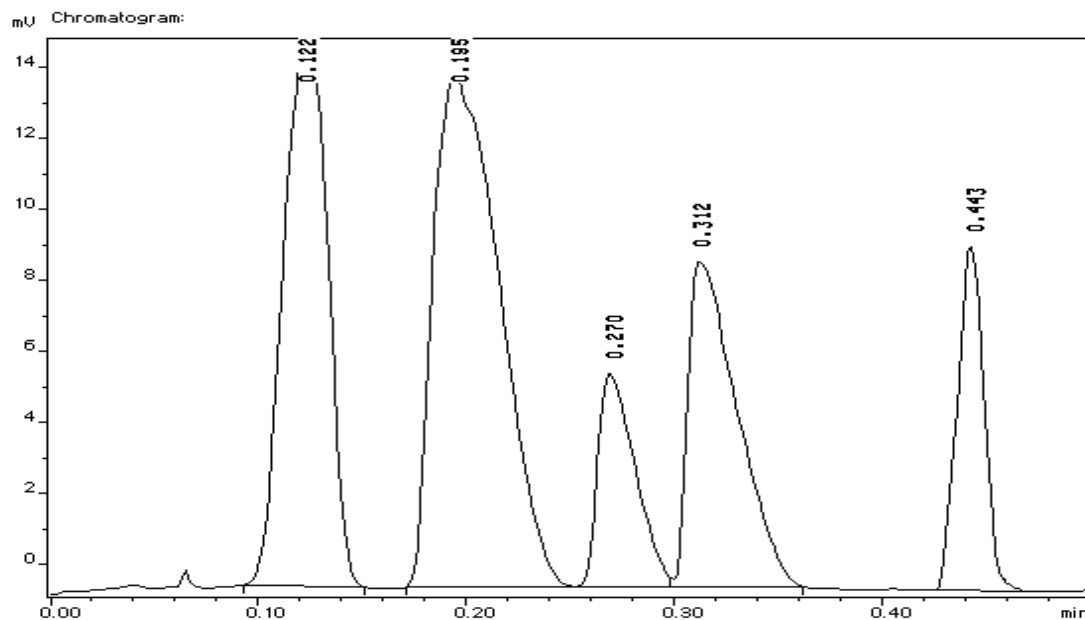


Figure 70. Standard curve of palmitic acid

3. TLC-FID chromatogram of standard oil



Peak No	Name	Ret. Time (min)	Pk. Start (min)	Pk. End (min)	Area	Height (mV)	Area %
1	Triolein	0.123	0.093	0.152	12059	15.26	26.731
2	Oleic acid	0.195	0.172	0.248	16658	14.41	36.923
3	1,3-Diolein	0.270	0.248	0.298	3913	5.99	8.674
4	1,2-Dioleoyl-rac-glycerol	0.313	0.298	0.362	7831	9.07	17.358
5	1-Monopalmitin	0.443	0.427	0.472	4654	9.67	10.315
					45115	54.39	100.000

Condition:

Stationary phase: CHROMAROD-SIII

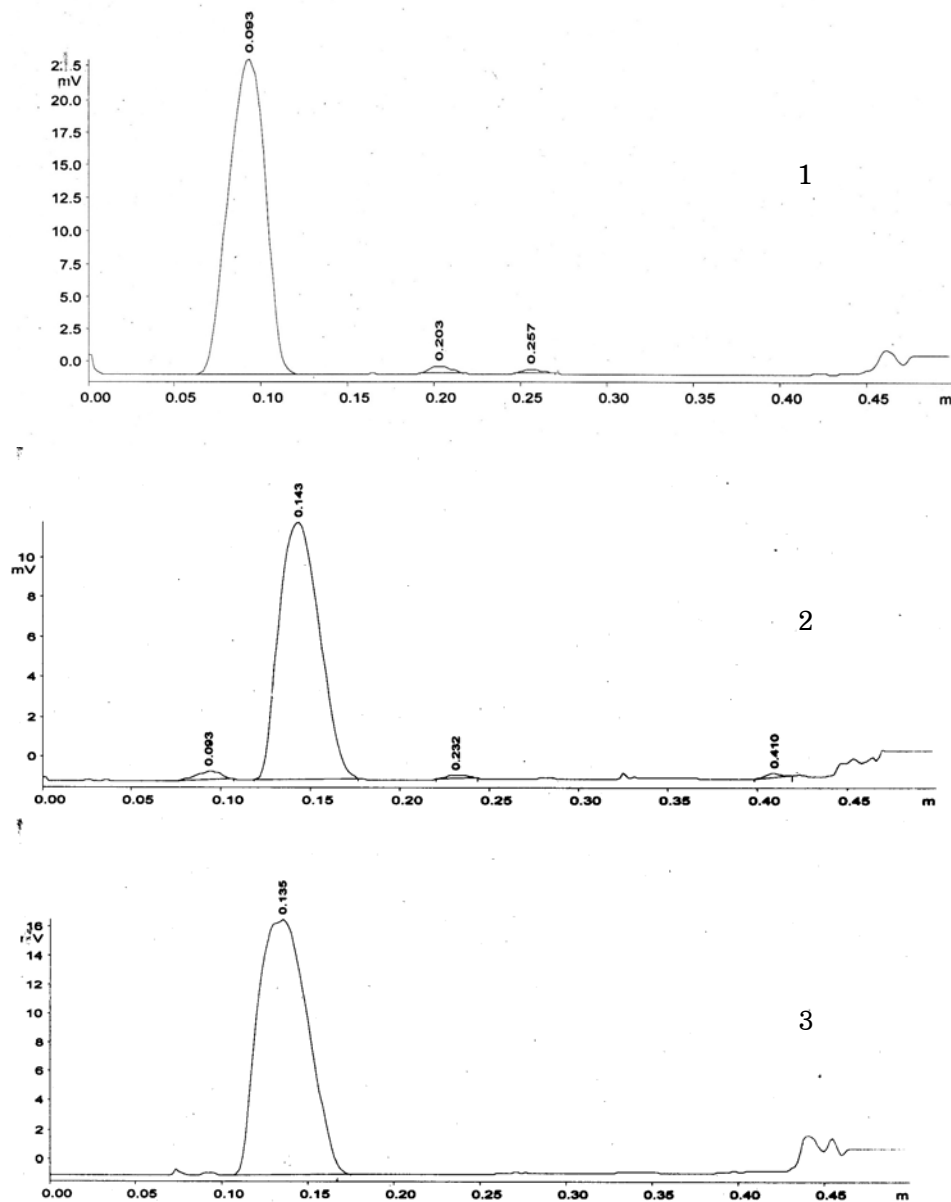
Mobile phase: benzene/chloroform/acetic acid (50:20:0.7)

Gas flow: H₂ 150 mL/min, air 700 mL/min

Scanning speed: 30 sec/Rod

Figure 71. TLC-FID chromatogram of standard oil.

4. TLC-FID chromatogram of palm oil and PFAD

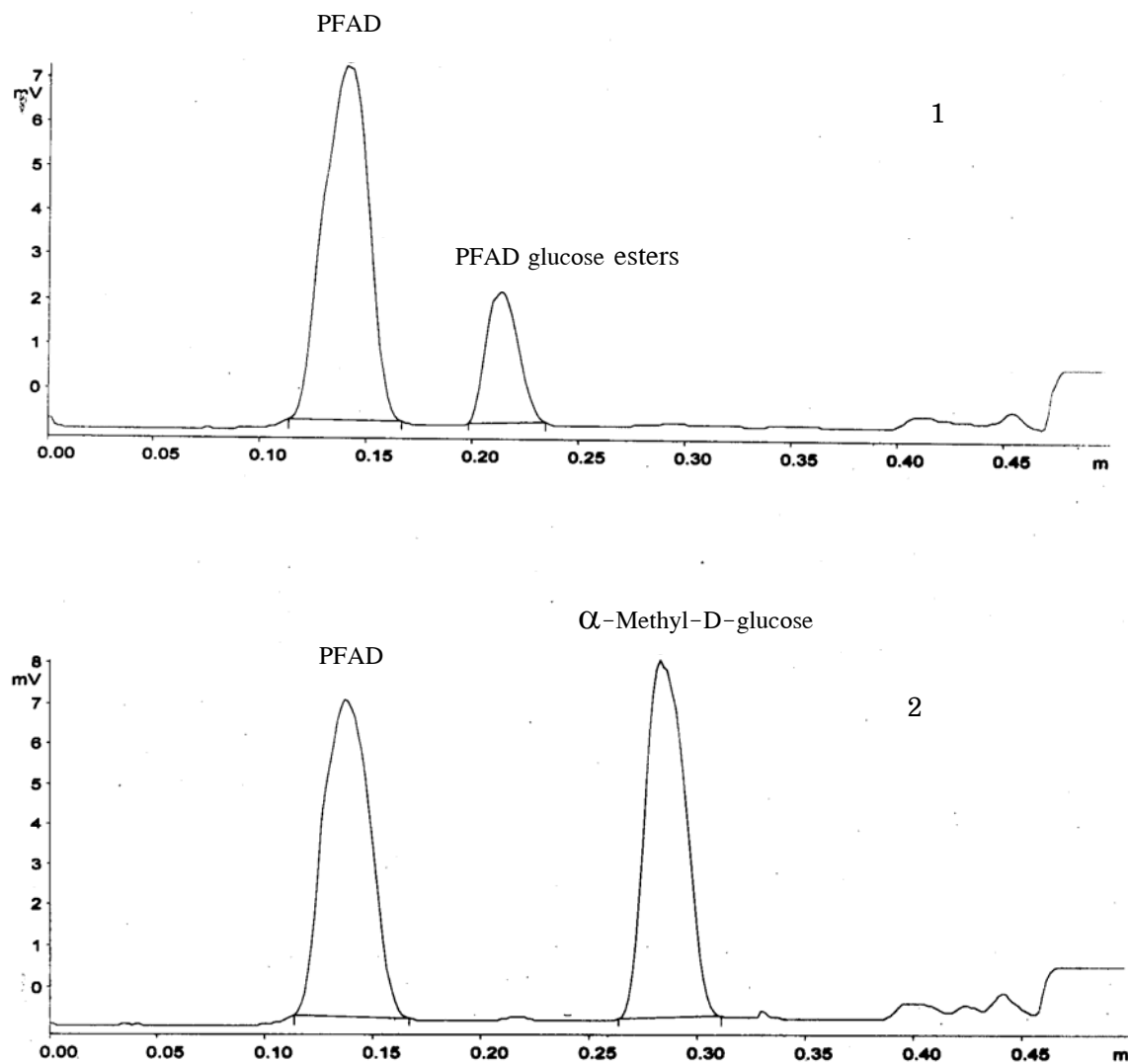


Condition:

Stationary phase: CHROMAROD-SIII
Mobile phase: Benzene/chloroform/acetic acid (50:20:0.7 v/v/v)
Gas flow: H₂ 150 mL/min, air 700 mL/min
Scanning speed: 30 sec/Rod

Figure 72. TLC-FID chromatogram of palm oil (1), crude PFAD (2) and partial purified PFAD (3).

5. TLC-FID chromatogram of PFAD glucose esters

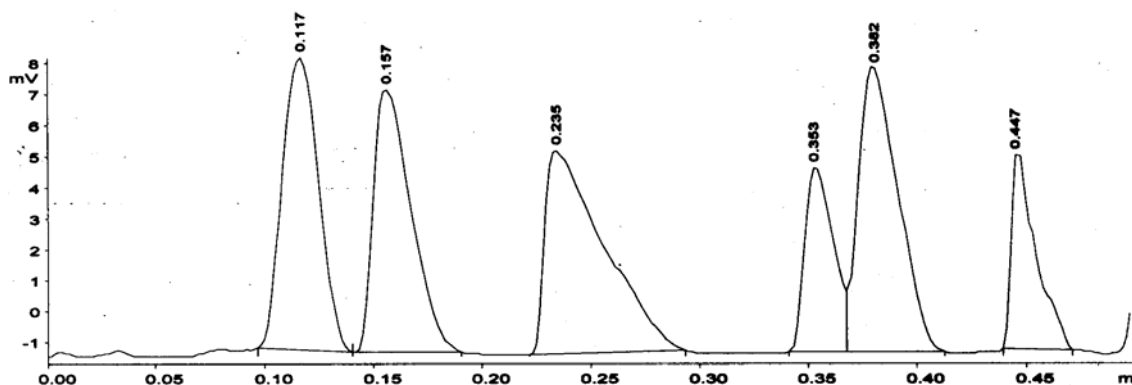


Condition:

Stationary phase: CHROMAROD-SIII
Mobile phase: Chloroform/methanol/formic acid (50:10:1 v/v/v)
Gas flow: H_2 150 mL/min, air 700 mL/min
Scanning speed: 30 sec/Rod

Figure 73. TLC-FID chromatogram of reaction mixture of PFAD glucose esters (1) synthesis and α -Methyl-D-glucose as standard sugar ester (2).

6. TLC-FID chromatogram of fatty acid methyl esters (FAME)



Peak No	Name	Ret. Time (min)	Pk. Start (min)	Pk. End (min)	Area	Height (mV)	Area %
1	Palmitic acid methyl ester	0.117	0.097	0.140	5792	9.43	19.790
2	Triolein	0.157	0.140	0.190	5339	8.49	18.245
3	Oleic acid	0.235	0.222	0.293	6462	6.58	22.082
4	1,3-Diolein	0.353	0.342	0.368	3022	5.99	10.327
5	1,2-Dioleoyl-rac-glycerol	0.382	0.368	0.413	6117	9.16	20.901
6	1-Monopalmitin	0.447	0.440	0.472	2533	6.31	8.655
					29265	45.95	100

Condition:

Stationary phase: CHROMAROD-SIII

Mobile phase: Chloroform/diethyl ether/formic acid (50:20:0.7) for 15 min
and benzene/hexane (50:50) for 35 minGas flow: H₂ 150 mL/min, air 700 mL/min

Scanning speed: 30 sec/Rod

Figure 74. TLC-FID chromatogram of standard oil compositions and FAME.

7. Determination of fatty acid compositions by GC analysis

Table 30. Retention time of standard fatty acid methyl esters.

Fatty acid	Retention time (min)
Caprylic acid (C8:0)	1.63–1.68
Caproic acid (C10:0)	2.50–2.53
Lauric acid (C12:0)	4.33–4.36
Myristic acid (C14:0)	7.10–7.17
Palmitoleic acid (C16:1)	9.50–9.51
Palmitic acid (C16:0)	9.75–9.76
Linolenic acid (C18:3)	11.97–11.98
Linoleic acid (C18:2)	12.05–12.06
Oleic acid (C18:1)	12.15–12.18
Stearic acid (C18:0)	12.56–12.57
EPA (C20:5)	14.99–15.00
DHA (C22:6)	21.90–21.91
Behenic acid (C20:0)	22.30

Column OPTIMA-5 (25 m x 0.25 mm i.d.)

Condition $T_1 = 150^\circ\text{C}$ (4°C/min, 0.50 min)

$T_2 = 170^\circ\text{C}$ (10°C/min)

$T_3 = 195^\circ\text{C}$ (10°C/min)

$T_4 = 215^\circ\text{C}$ (15 min)

Injection temperature 250°C

Detection FID (250°C)

Carrier gas Helium (1.24 mL/min)

8. Protein determination by Bradford's method (Bradford, 1976)

Protein content of enzymes was determined by Bradford's method.

The Bradford reagent is prepared by 100 mg of comassie brilliant blue G 250 is dissolved in 100 mL ethanol. Then 100 mL of 85% phosphoric is added and mixed well. The volume is adjusted to 600 mL by distilled water and then mixed with 100 mL glycerol. The volume is adjusted to 1 L by distilled water and undissolved substrate is filtered out. The Bradford solution is kept overnight at 4°C before use.

To determine of protein concentration, 100 mL of sample is mixed with 5.0 mL Bradford's reagent. This solution is allowed to stand for 5 min and the absorbance is measured at 595 nm. A standard curve is prepared using the protein under study or some other protein, such as serum albumin at concentrations of 100–1000 µg/mL.

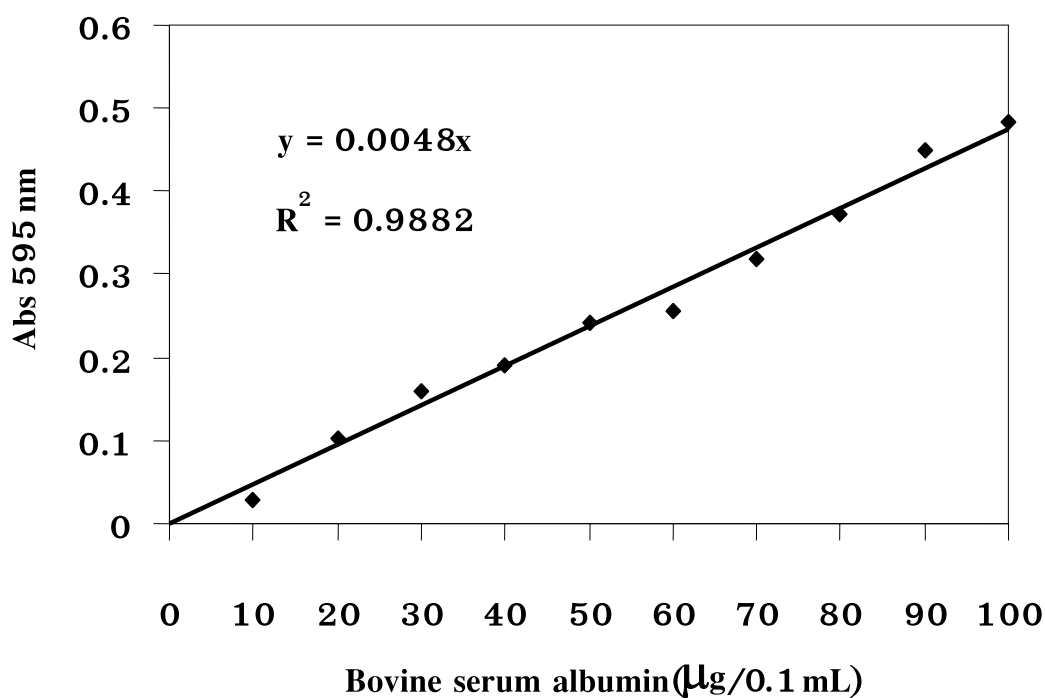


Figure 75. Standard curve of bovine serum albumin.

9. Saponification value (AOAC, 1999)**A. Reagent**

1. 0.1 N Alcoholic potassium hydroxide solution
2. 0.5 N HCl
3. 1% phenolphthalein

B. Determination

Accurately weight 5 g filtered sample into 250–300 mL Erlenmeyer flask. Pipette 50 mL alcoholic KOH solution into flask. Connect flask with air condenser and boil until fat is completely saponified (30 min). Cool and titrate with 0.5 N HCl using phenolphthalein as indicator. Conduct blank determination along with that on sample, using same pipette for measuring KOH solution and draining at the same time.

C. Calculation

$$\text{Calculate saponification number} = 28.05 (B-S)/g \text{ sample}$$

(mg KOH required to saponify 1 g fat or oil)

Where

$$B = \text{mL of 0.5 N HCl required by blank}$$
$$S = \text{mL of 0.5 N HCl required by sample}$$

10. Water activity determination

Water activity will be, by definition, equal in all phases at equilibrium. It is defined in any mixture as the ratio of the saturated vapor pressure of water present (p_w) to that of pure water (p_w°) at the same temperature (see equation below).

$$a_w = p_w / p_w^\circ$$

In this way, water activity is replaced by the equilibrium relative humidity. The fixed water activity of the reaction medium was determined by pre-equilibration of the reaction mixtures using saturated salt solution. To adjust the water activity, the immobilized enzymes or substrate solutions were equilibrated over saturated salt solutions in close vessels at room temperature for 72 hours. Salt solution with different water activity was used for this purpose (Table 31).

Table 31. Water activity of different saturated salt solutions at 25°C.

Saturated salt solution	Water activity (a_w)
LiBr	0.07
LiCl	0.11
CH ₃ COOK	0.25
MgCl ₂	0.33
K ₂ CO ₃	0.43
Mg(NO ₃) ₂	0.55
NaCl	0.75
K ₂ Cr ₂ O ₇	0.97

Source : Humeau *et al.* (1998)

11. Determination of molecular mass of protein by SDS-PAGE

The molecular mass of sample protein was determined by comparison the R_f of sample protein with the curve plotted between R_f of standard protein against their log molecular masses (log MW) under SDS-PAGE as below. The standard proteins are myosin (201 kDa), β -galactosidase (120 kDa), bovine serum albumin (100 kDa), ovalbumin (60 kDa), carbonic anhydrase (38 kDa), Soybean trypsin inhibitor (29.7 kDa), lysozyme and (20.7 kDa).

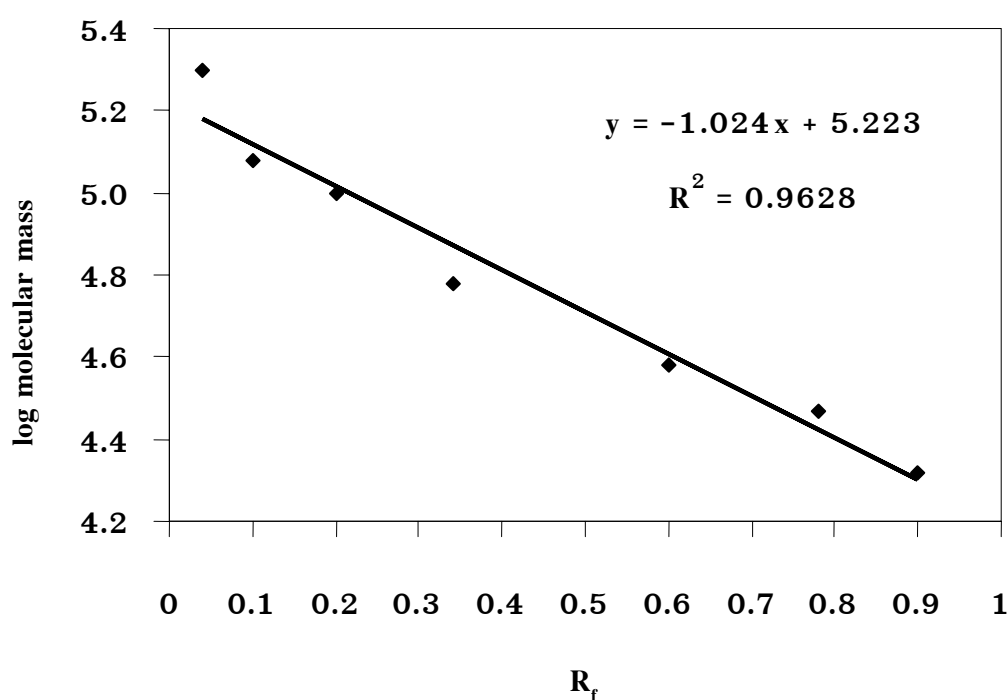
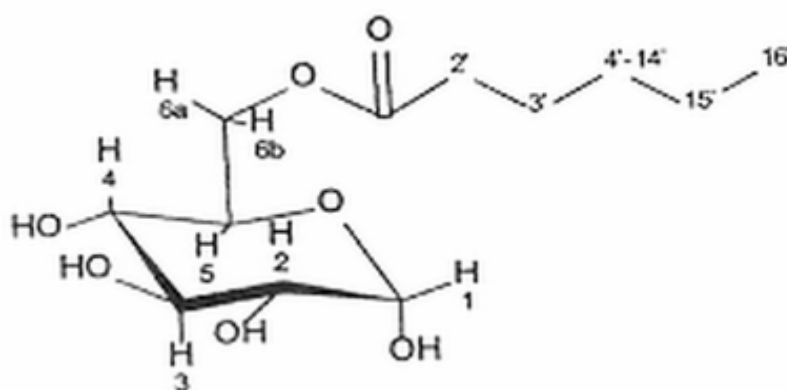


Figure 76. The curve of log molecular mass of standard protein against R_f under SDS-PAGE.

12. ^1H chemical shifts for 6-*O*-palmityl- α -D-glucopyranoside

Table 32. ^1H chemical shifts for 6-*O*-palmityl- α -D-glucopyranoside.

Proton position	C16- <i>O</i> -Glu ^a Chemical shift (ppm)	Glucose Chemical shift (ppm)
H-1	5.10	5.19
H-2	3.37	3.51
H-3	3.67	3.71
H-4	3.29	3.39
H-5	3.96	3.82
H-6a	4.35	3.82
H-6b	4.22	3.73
H-2'	3.32	
H-3'	1.61	
H-4'-H14'	1.27	
H-15'	1.30	
H-16'	0.87	



^aThe signals were relative to residual solvent signal: CHD_2OD 3.31 ppm run in $\text{CD}_3\text{OD}/\text{CDCl}_3$ (60:40) at 50°C.

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14. Nucleotide sequence of 16S rDNA gene of *Burkholderia multivorans* PSU-AH130

Sample Name: PSU-AH130

Identify : *Burkholderia multivorans*

16S rDNA Sequence (1490 bp)

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Blast result

Reference: Altschul, Stephen F., Thomas L. Madden, Alejandro A. Schäffer, Jinghui Zhang, Zheng Zhang, Webb Miller, and David J. Lipman (1997), "Gapped BLAST and PSI-BLAST: a new generation of protein database search programs", *Nucleic Acids Res.* 25:3389-3402.

RID: 1171300927-15257-136494918845.BLASTQ1

Database: All GenBank+EMBL+DDBJ+PDB sequences (but no EST, STS, GSS, environmental samples or phase 0, 1 or 2 HTGS sequences) 4,950,527 sequences; 19,726,293,847 total letters

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Length=1493

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Strand=Plus/Plus

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