

# Appendix

## The solutions

### 1. $\beta$ -Glycerophosphate 1 M, 50 ml

$\beta$ -Glycerophosphate	10.8 g
Sterile distilled water	50 ml
Sterile filter	

### 2. $\beta$ -Glycerophosphate 10 mM in culture medium

$\beta$ -Glycerophosphate 1 M	100 $\mu$ l
Culture medium.	10 ml
Aliquot 1000 $\mu$ l at -20°C	

### 3. $1\alpha,25$ -Dihydroxycholecalciferol (VD3) 100 nM stock solution, 240 $\mu$ l

VD3	10 $\mu$ g
Absolute ethanol	240 $\mu$ l

### 4. $1\alpha,25$ -Dihydroxycholecalciferol (VD3) 10nM in culture medium

VD3 100 nM	1 $\mu$ l
Culture medium	10 ml

**5. 2-Amino-2-methyl-1-propanol 0.75 mM, 1 L**

2-Amino-2-methyl-1-propanol 7.19 g

Deionized water 1 L

Adjust pH to 10.3 using 1M NaOH

**6. Acetic acid 2.5N /10M urea, 250ml**

Urea 150.2 g

Deionized water 100 ml.

Glacial acetic acid 35.9 ml

Adjust final volume to 250 ml

**7. Alcian blue solution, 100 ml**

Alcian blue 1.0 g

Glacial acetic acid 3% 100 ml

**8. Antibody diluting solution (1% Bovine serum albumin and 0.1% Sodium nitrite), 10 ml**

Bovine albumin 1 g

Sodium nitrite ( $\text{NaN}_3$ ) 0.1 g

DPBS 10 ml

**9. Ascorbic acid 5 mg/ml, 50 ml**

Ascorbic acid 0.25 g

Deionized water 50 ml

Sterile filtration

Aliquot 1000  $\mu\text{l}$  at  $-20^\circ\text{C}$

**10. Ascorbic 50  $\mu\text{g}/\text{ml}$  in culture medium**

Ascorbic acid 5 mg/ml	100 $\mu\text{l}$
Culture medium	10 ml of culture medium.

**11. Bovine thrombin 5000 Unit/ml stock solution, 2 ml**

Bovine thrombin	1000 Unit
Distilled water	2 ml
Aliquot 200 $\mu\text{l}$ and store at $-20^{\circ}\text{C}$	

**12.  $\text{CaCl}_2$  20% stock solution, 100 ml**

$\text{CaCl}_2$	20 g
Distilled water.	100 ml
Sterile filtered using 0.2 $\mu\text{m}$ pore filter	

**13. Chloroform/Methanol, 3:1, Solution, 4 L**

Chloroform	3 L
Methanol	1L
Mix well under chemical hood	

**14. Dexamethasone 500 $\mu\text{M}$  (200 ng/ $\mu\text{l}$ ) stock solution, 5 ml**

Dexamethasone	1 mg
Absolute ethanol	5 ml

**15. Dexamethasone 20 nM in culture medium**

Dexamethasone 500 $\mu\text{M}$	2 $\mu\text{l}$
Culture medium	50 ml

**16. EDTA 0.5M pH 8.0, 50 ml**

NaOH 10 N	50 ml
EDTA	186.12 g (0.5 mol)

Under hood with continuous stirring, Adjust pH to 8.0 using 30% HCL

**17. Electrophoresis gel loading buffer, 10 ml**

Glycerin	3.73 ml (50% v/v)
Xylencyanol	25.0 mg (0.25% w/v)
Bromphenolblau	25.0 mg (0.25% w/v)
Distilled water	6.27 ml

Sterile filtered

Store at -20°C for long term storage and at 4 °C for current usage

**18. Electrophoresis 0.2% gel, 300 ml**

Agarose	6 g
Tris-acetic-EDTA buffer (TAE) x 1	300 ml

Boiling in microwave at high level

**19. Ethidiumbromide 0.001%, 200 ml**

Ethidiumbromide	1 %
Deionized water	200 ml

**20. GuHCL 4M /50mM Tris-HCL-solution, 4 L**

GuHCL	1528.48 g
Tris	24.24 g
Deionized water	3.5 L

Continuous stirring until dissolve

Adjust volume to 4.0 L and Adjust pH to 7.4 with 25% HCL.

**21. Lysis buffer in ALP activity analysis**

2-Amino-2-methyl-1-propanol 0.75 mM	1ml
p-Nitrophenylphosphate (add Just before use add)	2 mg

Vortex briefly for mixing

**22. Masson-Trichrom-Goldner staining****Weigerts Eisenhaematoxylin**

Weigerts solution A and solution B	1:1
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**Masson solution (Goldner I)**

Solution A	33 ml
Solution B	66 ml
Solution C	20 ml, mix well
Acetic acid 0.2%	880 ml

**Solution A:**

Acidfuchsin	1 g
Acetic acid	1 ml
Deionized water	100 ml

**Solution B:**

Ponceau Xylidin	1 g
Acetic acid 100%	1 ml
Deionized water	100 ml

**Solution C:**

Azophloxin	0.5 g
Acetic acid 100%	0.2 ml
Deionized water	100 ml

**Orange-G solution**

Orange G	10 g
Deionized water	500 ml
Continuous stirring until dissolve, then add	
Molybdato-phosphoric acid	15 g

**Light green**

Light green	0.5 g
Acetic acid 100%	1 ml
Deionized water	500 ml

**23. Mineralized culture medium**

DMEM-F12	500 ml
Fetal bovine serum (FBS) (10%)	50 ml
Penicillin / Streptomycin solution (1%)	1 ml
Fungizone (0.1%)	0.5 ml
$\beta$ -Glycerophosphate	10 mM
Ascorbic acid	50 $\mu$ g/ml

**24. NaOH 10N, 100 ml**

NaOH pellet	4 g
Deionized water	100 ml
Continuous stirring using magnetic bar in plastic container	

**25. NaOH 2.7 N /1M HEPES, 250 ml**

NaOH 10 N	67.5 ml
HEPES	59.5 g
Deionized water	140 ml

Continuous stirring and Adjust volume to 250 ml using deionized water

**26. Neutral Buffered Formalin 10%, 1 L**

Sodiumphosphate, monobasic	4 g
Sodiumphosphate, dibasic	6.5 g
Formaldehyde 37%	100 ml
Continuous stirring	
Adjust pH to 6.8-7.0 with 10M NaOH	

**27. Neutral red solution 0.005% in culture medium**

Neutral red solution 0.5%	100 µl (sterile filtered)
Culture medium	10 ml
Mix well	

**28. Nuclear Fast Red**

Aluminium sulfate hydrated	25.0 g
Nuclear fast red	0.5 g
Deionized water	500 ml
Continuous stirring using magnetic bar	

**29. Oil Red O solution,**

Oil Red O	0.7 g
Propylene glycol	100 ml
Continuous stirring at 70°C	

**30. p-Nitrophenol solution 10  $\mu$ M – 250  $\mu$ M (Standard solution for ALP activity measurement) 1 L**

p-Nitrophenol	10-250 $\mu$ mol
Buffer for standard solution	1 L
Glycin	100 mM (7.507 g/L)
ZnCl <sub>2</sub>	1mM (0.13639 g/L)
MgCl <sub>2</sub>	1 mM (0.2033 g/L)
Deionized water	
Adjust pH to 9.6	
P-Nitrophenol solution 10 $\mu$ M – 250 $\mu$ M	

**31. Poly D-Lysin 1 mg/ml stock solution, 10 ml**

Poly-D-Lysin	10 mg
DPBS	10 ml
Store at -20°C	

**32. Propylene glycol 85%, 100 ml**

Propylene glycol	85 ml
Deionized water	15 ml

**33. Silver nitrate solution 5%, 100 ml**

Silver nitrate	5 g
Distilled water	100 ml

**34. Sodiumthiosulfate 5%**

Natriumthiosulfate	5 g
Deionised water	100 ml



**35. Thrombin 1000 Unit in 10% CaCl<sub>2</sub> solution, 100 µl**

Bovine thrombin 5000 Unit/ml	10 µl
CaCl <sub>2</sub> 20%	50 µl
Distilled water	40 µl

**36. Toluidine blue staining solutions (Solution A and Solution B)****Solution A:**

Di-sodium tetraborate	8 g
Toluidine blue	8 g
Deionized water	800 ml
Stirring for 15 minutes	

**Solution B:**

Pyronin G	2 g
Deionized water	200 ml
Stirring for 15 minutes	

**37. Tris-Acetic acid-EDTA (TAE ) x50 buffer, 500 ml**

Tris	121 g
Acetic acid 100%	28.55 ml
EDTA 0.5M	50 ml
Continuous stirring for complete dissolving	
Adjust volume to 500 ml using deionized water	

**38. Tris buffer saline (0.5 M), x10, 1 L**

NaCl	87.8 g
Tris	60.6 g
Deionized water	1 L
Adjust pH to 7.5 using 10% HCL.	

**39. Triton in PBS 0.2% solution, 1 ml**

Triton X-100

2  $\mu$ l

DPBS

1000  $\mu$ l

## VITAE

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### **Educational Attainment**

<b>Degree</b>	<b>Name of Institution</b>	<b>Year of Graduation</b>
Doctor of Dental Surgery (DDS)	Chulalongkorn University	1989
Master of Dental Science (MDS)	The University of Melbourne, Australia	1996
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### **Scholarship Awards during Enrolment**

	<b>Date</b>
German Academic Exchange Service (DAAD) (Codenummer: A/99/16805)	March 1999 – October 2001

### **Work-Position and Address:**

<b>Business Address</b>	<b>Date</b>	<b>Position</b>
Oral and Maxillofacial Surgery Department, Faculty of Dentistry, Prince of Songkla University, SONGKHLA	1989	Lecturer
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