# **APPENDIX** A

#### Method for the determination of inulin concentration (Davidson et al., 1963)

#### **Preparation of anthrone reagent**

- Add 100 ml of H<sub>2</sub>SO<sub>4</sub> slowly to 26 ml of ice-cooled DW in a flask immersed in an ice-bath. The mixture was allowed to cool.
- 2. Add 147 mg of anthron.
- 3. Stir until the mixture completely dissolved.

### Precipitation of plasma protein

- 1. Mix 30 µl of plasma with 1.8 ml of 3% trichloroacetic acid (CCl<sub>3</sub>COOH).
- 2. Mix well and stand for 10 min, shaking again at least once.
- 3. Centrifuge at 4000 rpm for 20 min to precipitated protein.

- Add 0.5 ml of DW (blank), fructose standard solution, plasma filtrate and diluted urine into cold test tubes, which the bottom of tubes are completed immersed in ice-bath. All determinations were done is duplicate.
- Add 3.0 ml of anthrone reagent, mix well. Cool in ice-bath before its place in water bath.
- 3. Place in water bath at 38°C for 50 min.
- 4. The concentration of inulin is determined directly from standard curve for range of 2 to 6.8 mg% of fructose.

## **APPENDIX B**

#### Method for the determination of para-aminohippuric acid concentration

(Smith et al., 1945)

#### **Solutions**

1. 0.1% sodium nitrite (NaNO<sub>2</sub>)

Add 0.1 g of NaNO<sub>2</sub> into DW and dilute to make 100 ml solution.

2. 0.5% ammonium sulfamate ( $H_6N_2O_3S$ )

Add 0.5 g of  $H_6N_2O_3S$  into DW and dilute to make 100 ml solution.

- 3. 0.1% N-(1-naphthyl)-ethylenediamine dihydrochloride ( $C_{12}H_{16}Cl_2N_2$ ) Add 0.1 g of  $C_{12}H_{16}Cl_2N_2$  into DW and dilute to make 100 ml solution.
- 4. 3.2% trichloroacetic acid (CCl<sub>3</sub>COOH)
  Add 3.2 g of CCl<sub>3</sub>COOH into distilled water and dilute to make 100 ml solution.
- 5. 0.2N hydrochloric acid (HCl)

Dilute 1.67 ml of HCl with DW, make up to 100 ml solution.

- Add 50 μl of PAH standard solution, plasma and diluted urine into 1100 μl of 3.2% CCl<sub>3</sub>COOH. Mix well and after about 10 min, centrifuge plasma for 2 min at 4000 rpm to precipitated protein. All determinations were done is duplicate.
- Pipette 1.0 ml of 3.2% CCl<sub>3</sub>COOH (blank) and solution from (1) into test tubes.

- 3. Add 0.2 ml of 0.2 N HCl and 0.1 ml of 0.1% NaNO<sub>2</sub>. Mix thoroughly, let stand not less than 3 min or more than 5 min after NaNO<sub>2</sub>.
- 4. Add 0.1 ml of 0.5%  $H_6N_2O_3S$ . Mix thoroughly, let stand not less than 3 min or more than 5 min later.
- 5. Add 0.1 ml of 0.1%  $C_{12}H_{16}Cl_2N_2$ , mix well.
- 6. Let stand at room temperature for 15-60 min.
- 7. Measure the developed color by the spectrophotometer at 540 nm.
- 8. The concentration of PAH is determined directly from standard curve for the range of 1-10 mg% of PAH.

# **APPENDIX C**

## Method for the determination of malondialdehyde content in renal tissue

#### homogenate (Modified from Ohkawa et al., 1979)

## **Solutions**

1. 8.1% sodium dodecyl sulfate ( $C_{12}H_{25}NaO_4S$ )

Add 8.1 g of  $C_{12}H_{25}NaO_4S$  into DW and dilute to make 100 ml solution.

2. 20% acetic acid

Dilute 20 ml acetic acid with DW and make up to 100 ml solution.

3. 0.8% thiobarbituric acid (TBA)

Add 0.8 g of TBA into DW, heat and stir until the solution completely dissolved. Make up to 100 ml solution.

4. 1.5 N sodium hydroxide (NaOH)

Add 6.122 g of NaOH into DW and dilute to make 100 ml solution.

5. n-butanol

- 1. Add 0.2 ml of DW, 1,1,3,3-tetramethoxypropane (MDA standard solution) and tissue homogenate solution into the test tubes for blank, standard and sample, respectively. All determinations were done is triplicate.
- Add 0.2 ml of 8.1% C<sub>12</sub>H<sub>25</sub>NaO<sub>4</sub>S and 1.5 ml of 20% acetic acid, mix thoroughly.
- 3. Adjust the pH of mixture to 3.5 with 1.5 N NaOH.
- 4. Add 1.5 ml of 0.8% TBA and make up to 4.0 ml with DW, mix well.

- 5. Heat for 60 min in 95°C water bath.
- 6. Cool in ice-bath.
- 7. Add 1.0 ml of DW and 5.0 ml of n-butanol, shake vigorously.
- 8. Centrifuge at 4000 rpm for 10 min.
- 9. Read absorbance of the organic layer (upper layer) at 532 nm.
- 10. The concentration of MDA is determined directly from standard curve of

1,1,3,3-tetramethoxypropane for the range of 0.2 to  $1 \times 10^{-4}$  M.

## **APPENDIX D**

# Method for the determination of protein content in renal tissue homogenate

(Itzhaki and Gill, 1964)

#### Preparation of biuret reagent (0.21% CuSO<sub>4</sub>·5H<sub>2</sub>O in 30% NaOH)

- 1. Add 0.525 g of CuSO<sub>4</sub>·5H<sub>2</sub>O into DW and dilute to make 50 ml solution.
- 75 g of NaOH is added to DW and diluted to make 200 ml solution.
   The solution was allowed to cool at room temperature.
- 3. Mix 50 ml of  $CuSO_4$ ·5H<sub>2</sub>O solution with 200 ml of NaOH solution.

- 1. Dilute tissue homogenate into 200 times with DW, mix thoroughly.
- Add 1 ml of DW, protein standard solution and tissue homogenate solution into test tubes for blank, standard and unknown, respectively.
   All determinations were done is triplicate.
- 3. Add to all tubes with 1 ml of biuret reagent, mix well.
- 4. Read absorbance at 310 nm after 10 min.
- The concentration of protein is determined directly from a standard curve of bovine serum albumin for the range of 50-650 μg.

## **APPENDIX E**

Effects of cisplatin on mean arterial blood pressure (MABP), urine flow rate ( $\dot{V}$ ), clearance of inulin ( $C_{in}$ ), clearance of para-aminohippuric acid ( $C_{PAH}$ ), sodium excretion rate ( $U_{Na} \dot{V}$ ), fractional excretion of sodium (FE<sub>Na</sub>), potassium excretion rate ( $U_K \dot{V}$ ), fractional excretion of potassium (FE<sub>K</sub>), plasma concentration of inulin and para-aminohippuric acid ( $P_{in}$  and  $P_{PAH}$ ) and blood urea nitrogen (BUN) in rats.

	vehicle	Cisplatin concentration (mg/kg bw)				
	( <b>n=6</b> )	4.5 (n=6)	6.0 (n=6)	7.5 (n=5)	9.0 (n=5)	
MABP (mmHg)	126 ± 9	$114 \pm 3$	$128 \pm 11$	$106 \pm 9$	$106 \pm 3$	
V (μl/min/g kw)	$13.58\pm2.85$	$21.55 \pm 3.54$	$15.88 \pm 3.40$	$30.06 \pm 5.40^{*\ddagger}$	$39.69 \pm 2.58^{*^{\ddagger}}$	
C <sub>in</sub> (ml/min/g kw)	$1.13\pm0.04$	$0.95\pm0.12$	$0.89\pm0.19$	$0.44 \pm 0.08^{*^{\ddagger}}$	$0.25 \pm 0.05^{*11}$	
C <sub>PAH</sub> (ml/min/g kw)	$3.82\pm0.13$	$3.24\pm0.68$	$2.41\pm0.62$	$0.51 \pm 0.20^{*^{\ddagger}}$	$0.22 \pm 0.05^{*^{\ddagger}}$	
U <sub>Na</sub> (mmol/min/g kw)	$2.32\pm0.58$	$3.48\pm0.53$	$2.27\pm0.47$	$3.15\pm0.43$	$4.01\pm0.35$	
FE <sub>Na</sub> (%)	$1.49\pm0.38$	$2.82\pm0.58$	$2.60\pm0.82$	6.48 ± 1.18*	$14.07 \pm 2.23^{*^{\ddagger}\#}$	
$U_{K} \dot{V}$ (mmol/min/g kw)	$0.56\pm0.06$	$0.66\pm0.09$	$0.50\pm0.10$	$0.23 \pm 0.04^{*^{\ddagger}}$	$0.16 \pm 0.02^{*^{\ddagger}}$	
FE <sub>K</sub> (%)	$14.46 \pm 1.35$	$19.32\pm0.83$	18.97 ± 2.73	$16.71 \pm 2.16$	$22.40\pm2.28$	
P <sub>in</sub> (mg%)	$21.27\pm0.77$	$28.87 \pm 3.20$	31.13 ± 7.68	38.40 ± 5.14	$60.48 \pm 6.48^{*^{\ddagger}\#}$	
P <sub>PAH</sub> (mg%)	$6.00\pm0.22$	$9.81 \pm 2.48$	$14.82\pm6.30$	$27.20 \pm 4.43^{*^{\ddagger}}$	38.82 ± 3.51* <sup>†‡</sup>	
BUN (mmol/l)	$2.9\pm0.2$	$4.2\pm1.0$	$5.2 \pm 1.6$	$11.7 \pm 1.8^{*^{\ddagger}}$	$16.1 \pm 1.1^{*^{\dagger^{\ddagger}\#}}$	

Animals were tested on the third day after cisplatin injection. Data are mean  $\pm$  S.E.M.

\*, <sup>†</sup>, <sup>‡</sup> and <sup>#</sup> P < 0.05 compared to vehicle and cisplatin treated group at the doses of 4.5, 6 and 7.5 mg/kg, respectively (one-way ANOVA with multiple comparison using Student-Newman Keuls post hoc test).

# **APPENDIX F**

Effects of *Hibiscus sabdariffa* Linn. extract on mean arterial blood pressure (MABP), urine flow rate ( $\dot{V}$ ), clearance of inulin ( $C_{in}$ ), clearance of para-aminohippuric acid ( $C_{PAH}$ ) sodium excretion rate ( $U_{Na}\dot{V}$ ), fractional excretion of sodium (FE<sub>Na</sub>), potassium excretion rate ( $U_K\dot{V}$ ), fractional excretion of potassium (FE<sub>K</sub>), plasma concentration of inulin and para-aminohippuric acid ( $P_{in}$  and  $P_{PAH}$ ) and blood urea nitrogen (BUN) in cisplatin-induced ARF rats.

	Short term treatment of HSE				Long term treatment of HSE				
	DW (n=6)	HSE (n=6)	C+DW (n=5)	C+HSE (n=6)	DW (n=6)	HSE (n=7)	C+DW (n=5)	C+HSE (n=7)	
MABP (mmHg)	$126\pm9$	$125\pm4$	$106 \pm 9$	$126\pm4$	$112\pm7$	$114\pm 6$	$93 \pm 2$	$100\pm5$	
V (μl/min/g kw)	$13.58\pm2.85$	$15.72\pm2.72$	$30.06 \pm 5.40^{*^{\dagger}}$	$22.85 \pm 1.99$	$14.80\pm3.26$	$9.52 \pm 1.71$	$28.13 \pm 2.31^{*^{\dagger}}$	$26.03 \pm 4.90^{*^{\dagger}}$	
C <sub>in</sub> (ml/min/g kw)	$1.13\pm0.04$	$1.30\pm0.05$	$0.44 \pm 0.08^{*\dagger}$	$0.80 \pm 0.17^{*^{\ddagger}}$	$1.08\pm0.02$	$1.18\pm0.07$	$0.34 \pm 0.03^{*\dagger}$	$0.58\pm0.16^{\star\dagger}$	
C <sub>PAH</sub> (ml/min/g kw)	$3.82\pm0.13$	$4.00\pm0.22$	$0.51 \pm 0.20^{*\dagger}$	$2.16 \pm 0.74^{*^{\ddagger}}$	$3.56\pm0.80$	3.63 ± 0.21	$0.34 \pm 0.05^{*^{\dagger}}$	$1.29 \pm 0.60^{*\dagger}$	
$U_{Na} \dot{V} (mmol/min/g kw)$	$2.32\pm0.58$	$2.90\pm0.72$	$3.15\pm0.43$	$3.49\pm0.50$	$3.00\pm0.94$	$1.51\pm0.27$	$3.88\pm0.68$	$3.40\pm0.46$	
FE <sub>Na</sub> (%)	$1.49\pm0.38$	$1.77\pm0.54$	$6.48 \pm 1.18^{*\dagger}$	$4.47 \pm 1.77$	$1.97\pm0.59$	$0.90\pm0.16$	$7.85 \pm 1.16^{*\dagger}$	$7.09 \pm 1.62^{*\dagger}$	
$U_K \overset{.}{V}$ (mmol/min/g kw)	$0.56\pm0.06$	$0.81\pm0.07$	$0.23 \pm 0.04^{*\dagger}$	$0.55\pm0.12^\ddagger$	$0.69\pm0.07$	$0.60\pm0.09$	$0.22\pm0.01^{\bigstar\dagger}$	$0.30\pm0.07^{\bigstar\dagger}$	
FE <sub>K</sub> (%)	$14.46\pm1.35$	$17.74 \pm 1.98$	$16.71\pm2.16$	$20.94 \pm 1.35$	$16.66 \pm 1.63$	$13.52\pm1.21$	$19.16 \pm 1.80$	$16.64 \pm 1.86$	
P <sub>in</sub> (mg%)	$21.27\pm0.77$	$22.52\pm0.56$	$38.40 \pm 5.14^{*^{\dagger}}$	$35.96 \pm 6.61^{*\dagger}$	$24.61\pm0.97$	$22.75\pm0.80$	$50.97 \pm 3.09^{*\dagger}$	$41.70 \pm 5.95^{*\dagger}$	
P <sub>PAH</sub> (mg%)	$6.00\pm0.22$	$7.26\pm0.20$	$27.20 \pm 4.43^{*\dagger}$	$22.34 \pm 8.41$	$7.21\pm0.23$	$6.95\pm0.49$	$37.45 \pm 3.26^{*\dagger}$	$25.94 \pm 5.69^{*^{\ddagger}}$	
BUN (mmol/l)	$2.9\pm0.2$	$2.5\pm0.1$	$11.7 \pm 1.8^{*^{\dagger}}$	$7.0 \pm 2.4^{\ddagger}$	$2.8\pm0.2$	$3.1\pm0.1$	$14.2 \pm 1.5^{*\dagger}$	$10.0 \pm 2.1^{*^{\ddagger}}$	

Animals were tested on the third day after cisplatin injection. DW = distilled water, HSE = Hibiscus sabdariffa Linn. water extract 250 mg/kg, C+DW = cisplatin 7.5 mg/kg + distilled water, C+HSE = cisplatin 7.5 mg/kg + Hibiscus sabdariffa Linn. water extract 250 mg/kg. Data are mean  $\pm$  S.E.M.

\*,  $\dagger$  and  $\ddagger P < 0.05$  compared with DW, HSE and C+DW groups, respectively (one-way ANOVA with multiple comparison using Student-Newman Keuls post hoc test).

# **APPENDIX G**

	vehicle (n=6)	Cisplatin concentration (mg/kg)					
		4.5 (n=10)	6.0 (n=6)	7.5 (n=12)	9.0 (n=9)		
MDA (nmol)	$6.29\pm0.45$	$7.67\pm0.33$	$7.58\pm0.68$	$8.32\pm0.30$	$7.36\pm0.34$		
Renal protein (mg)	$6.45\pm0.15$	$5.69\pm0.16$	$5.15\pm0.11$	$6.09\pm0.15$	$5.95\pm0.17$		
MDA (nmol/mg protein)	$0.97\pm0.05$	$1.35 \pm 0.06^{*}$	$1.47 \pm 0.14^{*}$	$1.38 \pm 0.06^{*}$	$1.25\pm0.08^{\bigstar}$		

Table 1 Effect of cisplatin on renal MDA level in rats.

Animals were tested on the third day after cisplatin injection.

Data are mean  $\pm$  S.E.M. \* P < 0.05 compared to vehicle group (one-way ANOVA with multiple comparison using Student-Newman Keuls post hoc test).

	Short term treatment of HSE				Long term treatment of HSE			
	DW (n=6)	HSE (n=8)	C+DW (n=12)	C+HSE (n=7)	DW (n=16)	HSE (n=19)	C+DW (n=6)	C+HSE (n=16)
MDA (nmol)	$6.29\pm0.45$	$7.02\pm0.32$	$8.32\pm0.30$	$5.51\pm0.40$	$7.36\pm0.34$	$7.58\pm0.68$	$8.32\pm0.30$	$7.36\pm0.34$
Renal protein (mg)	$6.45\pm0.15$	$6.64\pm0.13$	$6.09\pm0.15$	$6.07\pm0.15$	$5.95\pm0.17$	$5.15\pm0.11$	$6.09\pm0.15$	$5.95\pm0.17$
MDA (nmol/mg protein)	$0.97\pm0.05$	$1.06\pm0.04$	$1.38 \pm 0.06^{*\dagger}$	$0.91 \pm 0.08^{\ddagger}$	$1.14\pm0.06$	$1.55 \pm 0.04^{*}$	$1.44 \pm 0.12^{*}$	$1.45\pm0.09^{\bigstar}$

Animals were tested on the third day after cisplatin injection. DW = distilled water, HSE = Hibiscus sabdariffa Linn. water extract 250 mg/kg,

C+DW = cisplatin 7.5 mg/kg + distilled water, C+HSE = cisplatin 7.5 mg/kg + *Hibiscus sabdariffa* Linn. water extract 250 mg/kg.

Data are mean  $\pm$  S.E.M. \*, <sup>†</sup> and <sup>‡</sup> P < 0.05 compared with DW, HSE and C+DW groups, respectively (one-way ANOVA with multiple comparison

using Student-Newman Keuls post hoc test).